



СРПСКИ АРХИВ
ЗА ЦЕЛОКУПНО ЛЕКАРСТВО
SERBIAN ARCHIVES
OF MEDICINE

Address: 1 Kraljice Natalije Street, Belgrade 11000, Serbia
☎ +381 11 4092 776, Fax: +381 11 3348 653

E-mail: office@srpskiarhiv.rs, Web address: www.srpskiarhiv.rs

Paper Accepted*

ISSN Online 2406-0895

Original Article / Оригинални рад

Snježana Petrović¹, Jasmina Bašić^{2,†}, Zoran Mandinić³, Dragana D. Božić⁴,
Marina Milenković⁴, Zorica Vujić⁵

**Inhibitory effect of propafenone derivatives on *Pseudomonas aeruginosa*
biofilm and pyocyanin production**

Инхибиторни ефекат пропафенонских деривата на продукцију биофилма и
пиоцијанина код *Pseudomonas aeruginosa*

¹University of Belgrade, Institute for Medical Research, Centre of Research Excellence in Nutrition and Metabolism, Belgrade, Serbia;

²Medical College of Applied Sciences in Belgrade, Zemun, Serbia:

³University of Belgrade, School of Dental Medicine, Clinic for Pediatric and Preventive Dentistry, Belgrade, Serbia;

⁴University of Belgrade, Faculty of Pharmacy, Department of Microbiology and Immunology, Belgrade, Serbia;

⁵University of Belgrade, Faculty of Pharmacy, Department of Pharmaceutical Chemistry, Belgrade, Serbia

Received: July 27, 2018

Revised: July 11, 2019

Accepted: August 24, 2019

Online First: September 23, 2019

DOI: <https://doi.org/10.2298/SARH180727102P>

* **Accepted papers** are articles in press that have gone through due peer review process and have been accepted for publication by the Editorial Board of the *Serbian Archives of Medicine*. They have not yet been copy edited and/or formatted in the publication house style, and the text may be changed before the final publication.

Although accepted papers do not yet have all the accompanying bibliographic details available, they can already be cited using the year of online publication and the DOI, as follows: the author's last name and initial of the first name, article title, journal title, online first publication month and year, and the DOI; e.g.: Petrović P, Jovanović J. The title of the article. Srp Arh Celok Lek. Online First, February 2017.

When the final article is assigned to volumes/issues of the journal, the Article in Press version will be removed and the final version will appear in the associated published volumes/issues of the journal. The date the article was made available online first will be carried over.

†Correspondence to:

Jasmina BAŠIĆ

Medical College of Applied Sciences in Belgrade,

Cara Dušana 254, Zemun

Email: basic.jasmina23@gmail.com

Inhibitory effect of propafenone derivatives on *Pseudomonas aeruginosa* biofilm and pyocyanin production

Инхибиторни ефекат пропафенонских деривата на продукцију биофилма и пиоцијанина код *Pseudomonas aeruginosa*

SUMMARY

Introduction/Objective Biofilm and pyocyanin production are essential components of *Pseudomonas aeruginosa* virulence and antibiotic resistance. Our objective was to examine inhibitory effect of synthesized propafenone derivatives 3-(2-Fluorophenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride (5OF) and 3-(2-Trifluoromethyl-phenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride (5CF3) on biofilm and pyocyanin in *Pseudomonas aeruginosa* clinical strains.

Methods Effects were tested on nine clinical isolates and one control laboratory strain of *P. aeruginosa*. *In vitro* analysis of biofilm growing was performed by incubating bacteria (0.5 McFarland) with 5OF and 5CF3 (500 – 31.2 µg/ml) and measuring optical density at 570 nm. Bacteria in medium without compounds were positive control. Blank medium (an uninoculated medium without test compounds) was used as negative control. Pyocyanin production was estimated by optical density at 520 nm, after bacteria incubated with 5CF3 and 5OF (250 and 500 µg/ml), treated with chloroform, and chloroform layer mixed with HCl.

Results 500 µg/ml of 5OF and 5CF3 completely inhibited biofilm formation in 10/10 and 4/10 strains, respectively. 250 µg/ml of 5OF and 5CF3 strongly inhibited biofilm formation in 7/10 strains, while inhibition with 125 µg/ml of 5OF and 5CF3 was moderate. Lower concentrations had almost no effect on biofilm production. Pyocyanin production was reduced to less than 40% of the control value in 6/9, and less than 50% of the control in 7/9 strains with 500 µg/ml of 5OF and 5CF3, respectively. At 250 µg/ml 5OF and 5CF3 most strains had pyocyanin production above 50% of the control value.

Conclusion Synthesized Propafenone derivatives, 5OF and 5CF3, inhibited biofilms and pyocyanin production of *Pseudomonas aeruginosa* clinical strains. Presented results suggest that propafenone derivatives are potential lead-compounds for synthesis of novel antipseudomonal drugs.

Keywords: Propafenone derivatives; *Pseudomonas aeruginosa*; biofilm; pyocyanin

САЖЕТАК

Увод/циљ Производња биофилма и пиоцијанина су важни фактори вируленције и антибиотске резистенције бактерије *Pseudomonas aeruginosa*. Наш циљ био је испитати инхибиторни ефекат синтетисаних пропафенонских деривата, 3-(2-Флуоро-фенил)-1-[2-(2-хидрокси-3-пропиламино-пропокси)-фенил]-пропан-1-он-хидрохлорид (5OF) и 3-(2-Трифлуорометилфенил)-1-[2-(2-хидрокси-3-пропиламино-пропокси)-фенил]-пропан-1-он-хидрохлорид (5CF3), на продукцију биофилма и пиоцијанина код клиничких изолата *P. aeruginosa*.

Метод Ефекат пропафенонских деривата испитан је на девет клиничких изолата и једном стандардном соју *P. aeruginosa*. Утицај на продукцију биофилма испитан је *in vitro*, инкубацијом бактерија (0.5 по МекФарланд-у) са 5OF и 5CF3 (500–31.2 µg/ml), и мерењем оптичке густине на 570 nm. Бактерије у медијуму без испитиваних једињења су биле позитивна, а сам медијум негативна контрола. Производња пиоцијанина одређивана је мерењем оптичке густине на 520 nm, на конинкубације бактерија са 5CF3 или 5OF (250 и 500 µg/ml), третмана хлороформом и мешања хлороформског слоја са HCl.

Резултати При концентрацији од 500 µg/ml 5OF је довео до потпуне инхибиције продукције биофилма код свих испитиваних сојева (10/10). Инхибиција биофилма са 500 µg/ml 5CF3 била је потпуна код 4/10 сојева. При концентрацији 5OF и 5CF3 од 250 µg/ml продукција биофилма код већине испитаних изолата била је слаба, док је при концентрацији 125 µg/ml 5OF односно 5CF3 продукција била умерена. Ниже концентрације 5OF и 5CF3 нису имале инхибиторни ефекат на формирање биофилма. У присуству 500 µg/ml 5OF у 6/10 испитиваних сојева продукција пиоцијанина пала је на мање од 40% у односу на контролну вредност. Иста концентрација (500 µg/ml) 5CF3 снижила је продукцију пиоцијанина на мање од 50% од контроле у 7/9 сојева. При концентрацији 250 µg/ml 5OF или 5CF3 већина сојева продуковала је пиоцијанин изнад 50% у односу на позитивну контролу.

Закључак Синтетисани пропафенонски деривати, 5OF и 5CF3, инхибирају продукцију биофилма и пиоцијанина код клиничких сојева *P. aeruginosa*. Добијени резултати указују да пропафенонски деривати представљају могућа полазна једињења за синтезу нових антипсеудомонас агенаса.

Кључне речи: пропафенонски деривати; *Pseudomonas aeruginosa*; биофилм; пиоцијанин

INTRODUCTION

As an opportunistic human pathogen, *Pseudomonas aeruginosa* has evolved a number of immunoevasive strategies to impair host defense, including growing in biofilm [1, 2]. Biofilms are bacterial clusters encased in self-produced polymeric matrix attached to the epithelial surfaces or surface of medical implants. They are characterized by lower metabolic activity, increased synthesis of protective molecules, prolonged doubling time and genetic diversity of bacterial cells, all together improving bacterial tolerance to antibiotics and survival in harsh conditions [3, 4]. Biofilm production in *Pseudomonas aeruginosa* is well known causative agent of antibiotic resistant infections in humans, such as pneumonia, and infections in patients with bronchiectasis and cystic fibrosis [5, 6]. Due to resistance to phagocytosis and pronounced antibody response, those infections lead to chronic inflammation, often with severor fatal outcome [7, 8]. Thus, there is an urgent need to develop new drugs for the treatment of *Pseudomonas aeruginosa* biofilm-associated infections. In addition, *Pseudomonas aeruginosa* pathogenicity is intimately linked to its ability to produce large variety of virulence factors, including phenazines, and most abundant pyocyanin.[9, 10]. Pyocyanin is highly diffusible blue pigment, which can interact with molecular oxygen and stimulate generation of oxygen radicals, leading to redox disbalance, injury and death of host cells [11]. As virulence factor in chronic lung infection pyocyanin disrupts redox control, inhibits respiration in human cells, accelerates neutrophil apoptosis, therefore impairing host defense and favoring bacterial persistence [12, 13, 14].

Considering that ion channels are integral part of each living cell which play a key role in cell division, proliferation, excitation, and apoptosis, modulators of ion channel activity have become important target molecules in medical chemistry [15]. Propionophenone is relatively simple compound commercially obtained from benzoic and propionic acid, it has channel-modulatory effect and serve as a precursor of numerous drugs (e.g.ephedrine,

arylalkene) [16, 17, 18]. Propionophenone derivatives called propafenones are primarily known on their antiarrhythmic action, but they are also involved in treatment of many different diseases including lupus erythematosus, epilepsy, Alzheimer disease, malaria, ebola, cancer [19, 20,21, 22, 23, 24,25]. In addition, recent studies have shown that analogs of propafenone exhibit antifungal activity [26]. Therefore, the molecule of propafenone has become a model of compounds used in multidrug-resistant studies [27].

Since data on antibacterial activity of propafenone derivatives are scarce, we decided to test potential antibacterial activity of 3-(2-Fluoro-phenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride (5OF) and 3-(2-Trifluoromethyl-phenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride (5CF3). Even more, because the influence of propafenone derivatives on the *Pseudomonas aeruginosa* biofilm and pyocyanin production has not yet been tested, we expanded our examinations on the influence of propafenone derivatives on expression of *Pseudomonas aeruginosa* virulence factors.

In the present study, we aimed to evaluate the inhibitory effects of *ortho*-fluorinated propafenone derivatives, which were synthesized in our laboratory, on biofilm and pyocyanin production in *Pseudomonas aeruginosa* clinical strains.

METHODS

Effect of *ortho*-fluorinated propafenone derivatives on the *Pseudomonas aeruginosa* biofilms

Test compounds

Ortho-fluorinated propafenone derivatives were synthesized at the Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Belgrade, Serbia: 5CF3: 3-(2-Trifluoromethyl-phenyl) -1- (2- (2-hydroxy-3-propylamino-propoxy) -phenyl) -propan-1-

one hydrochloride and 5OF: 3- (2-Fluoro-phenyl) -1- (2- (2-hydroxy-3-propylamino-propoxy) -phenyl) -propan-1-one hydrochloride [28]. The structure of synthesized derivatives was spectrophotometrically analysed at FT-IR spectrophotometer Nicolet iS10 (Thermo Fisher Scientific Inc., SAD) [29].

The stock solutions of 5CF3 or 5OF (1 mg/ml) were prepared in 5% DMSO. The working solutions were prepared in trypticase soy bean broth with the addition of 1% glucose (TSB, Lab M Limited, UK) according to Knobloch et al. (2002) [30]. The concentrations of working solutions of 5CF3 or 5OF were 31.2, 62.5, 125, 250 and 500 µg/ml. In previous studies, we already investigated antimicrobial effect of tested compounds in the concentration range from 500 µg/ml to 62.5 µg/ml, and the best activity was observed for 250 µg/ml and 500 µg/ml of 5OF and 5CF3 [31].

***Pseudomonas aeruginosa* clinical isolates**

The effects of tested compounds were investigated on nine clinical isolates obtained from urine (strains 1, 2, 5, 8, 9), ear swab (strains 3, 6, 7) or sputum (strain 4) and one laboratory control strain (ATCC 27853). Bacteria were stored at -70°C in brain-heart infusion broth (Lab M Limited, UK) until needed.

Culture medium

Trypticase soy bean broth with 1% glucose (TSB) and trypticase soy bean agar (TSA) both from Lab M Limited, UK, were used.

Analysis

Biofilm production and quantification were performed according to protocols described by Stepanović et al. [32]. Briefly, bacteria were resuspended in saline to the

density of a 0.5 McFarland standard ($\sim 10^8$ CFU/ml). In 96 microtiter plates 180 μ l of test compounds and 20 μ l of bacterial suspension were added in triplicate. Bacteria incubated in medium without test compounds were used as positive control, while blank medium (uninoculated medium without test compounds) represented negative control. After incubation which lasted 24h at 35 °C, plates were washed in phosphate buffer (PBS, pH 7.2), dried, fixed with methanol and stained with 2% crystal violet (Himedia, India). After washing, the color was extracted from bacteria with 96% ethanol. The optical density (OD) was measured spectrophotometrically at 570 nm (ICN Flow Titertek Multiscan Plus, ICN, USA). Each experiment was repeated three times. To calculate the category of biofilm production, the optical density cut-off (ODc) was determined as three standard deviations above the mean OD of the negative control. According to the calculated results, all tested strains were categorized into four groups: $OD \leq ODc$ - category 0 (no biofilm production); $ODc < OD \leq 2xODc$ - category 1 (weak biofilm production); $2xODc < OD \leq 4xODc$ - category 2 (moderate biofilm production) and $4xODc < OD$ - category 3 (strong biofilm production).

Effect of *ortho*-fluorinated propafenone derivatives on the *Pseudomonas aeruginosa* pyocyanin production

Test compounds

Ortho-fluorinated propafenone derivatives, 5CF3 and 5OF, were synthesized at the Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Belgrade, Serbia.

Working dilutions of 250 and 500 μ g/ml 5CF3 and 5OF in 5% DMSO were prepared from the stock solution of 1 mg/ml in 5% DMSO. Working concentrations were chosen on

the basis of our results about 5CF3 and 5OF effect on biofilm formation, where concentrations of 250µg/ml and 500µg/ml appeared to have the strongest inhibitory effect.

***Pseudomonas aeruginosa* clinical strains**

The effects of tested compounds were investigated on nine *Pseudomonas aeruginosa* clinical isolates. The sources of bacteria and storage conditions were the same as previously described in section Methods.

Culture medium

Mueller-Hinton broth for bacteria (Torlak, Serbia) was used.

Pyocyanin determination

Pyocyanin was determined as previously described by Glamočlija et al. (2015) [33]. A 5 ml of bacterial cultures in exponential phase of growth were incubated with test compounds for 24h at 37 °C and then treated with 3 ml of chloroform. Separated chloroform layer was mixed with 1 ml of 0.2 M HCl. Optical density was measured at 520 nm [34]. Positive controls for each isolate were cultivated at the same conditions in medium without tested compounds. Values were expressed as a ratio $(OD_{520}/OD_{600}) \times 100$. Two experiments, each in triplicate, were performed. Results were calculated as the percent of the pyocyanin production compared to the positive control (expressed as $100\% \pm SD$).

Statistical analysis

Obtained data were analyzed using statistical analysis software package - SPSS Statistics (PASW statistics for Windows, Version 18.0, Chicago: SPSS Inc. USA) and Student's *t*-test [35].

RESULTS

Effect of *ortho*-fluorinated propafenone derivatives on the *Pseudomonas aeruginosa* biofilm formation

Both *ortho*-fluorinated propafenone derivatives, 5OF and 5CF3, inhibited production of *Pseudomonas aeruginosa* biofilms. The intensity of inhibitory effects changed in concentration dependent manner, thus, higher drug concentrations lead to stronger inhibition. The highest inhibition occurred at concentration of 500 µg/ml of both compounds. When the 5OF and 5CF3 concentrations decreased to 250 µg/ml, 125 µg/ml, 62.5 µg/ml or 31.2µg/ml the inhibitory effect was also decreased. In addition, there was a difference in different isolates sensitivity to particular drug concentration. 500 µg/ml of 5OF and 5CF3 completely inhibited biofilm formation in 10/10 and 4/10 strains, respectively. 250 µg/ml 5OF and 5CF3 strongly inhibited biofilm formation in 7/10 strains, while inhibition with 125 µg/ml 5OF and 5CF3 was moderate. In the presence of lower 5OF and 5CF3 concentrations, 62.5 µg/ml and 31.2, 8/10 tested strains exerted strong biofilm production. Categories of biofilm production in different isolates and in the presence of various concentrations of tested compounds are presented in Table 1 and 2.

Effect of *ortho*-fluorinated propafenone derivatives on the *Pseudomonas aeruginosa* pyocyanin production

Both *ortho*-fluorinated propafenone derivatives, 5CF3 and 5OF, inhibited production of pyocyanin in *Pseudomonas aeruginosa*. In the presence of 500 µg/ml 5OF or 5CF3 production of pyocyanin was reduced to less than 40% of the control value in 6/9 strains, and less than 50% of the control in 7/9 strains, respectively. In the presence of 250µg/ml 5OF or 5CF3 most strains had pyocyanin production above 50% of the control value. The difference in the sensitivity to the tested compounds among various strains was also detected. Results of

inhibitory action of 5OF and 5CF3 on the pyocyanin production in *Pseudomonas aeruginosa* are expressed as the percentage of the absorbance of positive controls (presented as 100% \pm SD) (Table 3).

DISCUSSION

Antibiotic compounds that inhibit different virulence factor, such as enterotoxins, hemolysins, biofilm or pigments, became a focus of recent research [36]. The resistance of *Pseudomonas aeruginosa* isolates to antimicrobial drugs is largely attributed to its ability to form a biofilm and produce bacterial pigment pyocyanin [37]. In this study we have used synthesized propafenone derivatives, 5CF3 and 5OF, to test inhibitory effect on *Pseudomonas aeruginosa* biofilm and pigment production.

Antimicrobials are generally dedicated to kill bacteria (bactericidal) or to inhibited bacterial growth (bacteriostatic). But, mostly due to frequent chromosomal mutations, *Pseudomonas aeruginosa* appeared to be extremely adaptive and acquired resistance to many antibiotics such as carbapenems, penicillins and cephalosporins. Recent efforts to develop novel class of anti-pseudomonas agents moved their focus to *Pseudomonas aeruginosa* physiology and collective behavior of bacterial population [38]. Therefore, biofilm formation and its modulation became a subject of our research interest. Our results have shown that propafenone derivative 5OF and 5CF3 significantly reduced biofilm production in all tested isolates of *Pseudomonas aeruginosa*. Previous study on propafenone compounds reported antimicrobial effect due to inhibition of ubiquitous bacterial multidrug efflux pumps [39]. Thus, by channels-blocking propafenone may decrease drug resistance and positively impact on clinical outcome of *Pseudomonas aeruginosa* infections [40]. On the other hand, to the best of our knowledge, this study revealed identification of *ortho*-fluorinated propafenone derivatives as efficient agents that inhibit *Pseudomonas aeruginosa* biofilm formation for the

first time. The inhibitory effects of both, 5OF and 5CF3, were found. Numerous external factors affect biofilm formation by *Pseudomonas aeruginosa*. Also, the type of tissue has strong impact on biofilm formation, and researchers commonly test biofilm formation of *Pseudomonas aeruginosa* from a variety of clinical sources [41]. In our study, various clinical strains showed differences in sensitivity to tested compounds, but those variations were not connected to specific bacterial source (urine, ear swab, sputum). However, we tested only nine clinical isolates (5 -urine, 3 - ear swab, 1 – sputum) and for such a small number of samples statistical data processing is not relevant.

The highest tested dose of both compounds (500 µg/ml) was the most efficient, reducing bacterial growth to the highest extent. However, when the concentration of test agents decreased bacterial growth recovered. In present study, 5OF was more effective in reducing bacterial growth compared to 5CF3. This could be explained by higher binding affinity to bacterial transport porin in a case of monofluorinated propafenone derivatives (such as 5OF), compared to trifluoromethyl derivative (5CF3), as found in our *docking* studies (data not shown) [31]. Namely, biofilm formation depends on the presence of an extracellular matrix which is mixture of polysaccharides, proteins, and nucleic acids (extracellular DNA). Matrix polysaccharides (alginate and lipopolysaccharides) which are synthesized in bacterial cytoplasm, bind to membrane transporters to be extruded out of the cell [42]. It was found that both fluorinated derivatives tested in this study briefly occupied key substrate-specific sites in the bacterial porin (Arg124). This discovery might be associated with interruption of the transport of carbohydrate compounds involved in synthesis of biofilm [43].

The blue pigment pyocyanin, chemical derivative of phenazine, is one of the most important virulence factors in *Pseudomonas aeruginosa* [44]. Pyocyanin is toxic for respiratory epithelium, it acts on the cell structure and function, disrupts normal expression of

genes involved in efflux pumps, redox homeostasis and iron acquisition in human cells [45, 46, 47]. Thus, control of pyocyanin production may be a mechanism to reduce bacterial pathogenicity. Results of our study have shown that both, 5OF and 5CF3, inhibited production of pyocyanin in all tested *Pseudomonas aeruginosa* isolates. The inhibitory effect was concentration dependent, higher concentrations caused stronger inhibition, while inhibitory effect decreased with lower drugs concentration. Literature survey on other drugs suggests that *ortho*-fluorinated propafenone derivative 5OF had significantly stronger inhibitory effect on the production of pyocyanin in *Pseudomonas aeruginosa* strains than commercial antibiotics ampicillin or streptomycin. Namely, we have shown that lower concentrations of 5OF, 500 µg/ml and 250 µg/ml, exerted same or even enhanced inhibitory effect compared to commercial antibiotics when applied in 2-4 fold higher concentration (1 mg/ml) [33, 48]. Similarly, the concentrations of 5CF3 which led to pyocyanin reduction were within the same range as concentration of standard drugs. Observed propafenone-induced pyocyanin inhibition could be discussed in a view of recent results on pyocyanin impact on extracellular DNA (eDNA) and biofilm formation [11]. It was found that pyocyanin decreases eDNA content within the *Pseudomonas aeruginosa* biofilm. Since eDNA promotes bacterial adhesion and cellular aggregation, depletion of eDNA can reduce biofilm strength and disturb protection of bacterial cells against antibiotics. Based on above, it was assumed that reduction of pyocyanin production as detected in our study could be a model of propafenone derivatives action against *Pseudomonas aeruginosa* pathogenicity and infection.

CONCLUSION

The results of the study suggest that synthesized *ortho*-fluorinated propafenone derivatives inhibit biofilm and pyocyanin production in *Pseudomonas aeruginosa* clinical

strains. Presented results suggest that propafenone derivatives could be considered as potential lead-compounds for synthesis of novel antipseudomonal drugs.

ACKNOWLEDGEMENT

This work was supported by Serbian Ministry of Education, Science and Technological Development [OI 172041] and [III 41030]. The authors declare no conflict of interest.

This work is a part of Jasmina Bašić's PhD thesis "Examination of correlation between chemical structure, physicochemical properties and retention parameters and antimicrobial activity of newly synthesized derivatives of propiophenone".

Conflict of interest: None declared.

REFERENCES

1. Hilker R, Munder A, Klockgether J, Losada PM, Chouvarine P, Cramer N, et al. Interclonal gradient of virulence in the *Pseudomonas aeruginosa* pangenome from disease and environment. *Environ Microbiol*. 2015; 17(1):29-46. [DOI: 10.1111/1462-2920.12606] [PMID: 25156090]
2. Streeter K, Katouli M. *Pseudomonas aeruginosa*: A review of their pathogenesis and prevalence in clinical settings and the environment. *Infect Epidemiol Med*. 2016; 2(1):25-32. [DOI: 10.18869/modares.iem.2.1.25]
3. De Kievit TR. (2009). Quorum sensing in *Pseudomonas aeruginosa* biofilms. *Environ Microbiol*. 2009; 11(2):279-88. [DOI:10.1111/j.1462-2920.2008.01792.x] [PMID:19196266]
4. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug resistant, extensively drug resistant and pandrug resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect*. 2012; 18(3):268-81. [DOI:10.1111/j.1469-0691.2011.03570.x] [PMID:21793988]
5. Gillis RJ, White KG, Choi KH, Wagner VE, Schweizer HP, Iglewski BH. Molecular basis of azithromycin-resistant *Pseudomonas aeruginosa* biofilms. *Antimicrob Agents Chemother*. 2005; 49(9):3858-67. [DOI:10.1128/AAC.49.9.3858-3867.2005] [PMID:16127063]
6. Johansson EM, Crusz SA, Kolomiets E, Buts L, Kadam RU, Cacciarini M, et al. Inhibition and dispersion of *Pseudomonas aeruginosa* biofilms by glycopeptide dendrimers targeting the fucose-specific lectin LecB. *Chem Biol*. 2008; 15(12):1249-57. [DOI:10.1016/j.chembiol.2008.10.009][PMID:19101469]
7. Kuang Z, Hao Y, Walling BE, Jeffries JL, Ohman DE, Lau GW. *Pseudomonas aeruginosa* elastase provides an escape from phagocytosis by degrading the pulmonary surfactant protein-A. *PLoS one*. 2011; 6(11):e27091 [DOI:10.1371/journal.pone.0027091][PMID:22069491]
8. Pressler T, Bohmova C, Conway S, Dumcius S, Hjelte L, Høiby N, et al. Chronic *Pseudomonas aeruginosa* infection definition: Euro Care CF working group report. *J Cyst Fibros*. 2011; 10:S75-S78. [DOI:10.1016/S1569-1993(11)60011-8] [PMID:21658646]
9. Potron A, Poirel L, Nordmann P. Emerging broad-spectrum resistance in *Pseudomonas aeruginosa* and *Acinetobacter baumannii*: mechanisms and epidemiology. *Int J Antimicrob Agents*. 2015; 45(6):568-85. [DOI:10.1016/j.ijantimicag] [PMID:25857949]
10. Winstanley C, O'Brien S, Brockhurst MA. *Pseudomonas aeruginosa* evolutionary adaptation and diversification in cystic fibrosis chronic lung infections. *Trends Microbiol*. 2016; 24(5):327-37. [DOI:10.1016/j.tim.2016.01.008] [PMID:26946977]
11. Das T, Manefield, M. Pyocyanin promotes extracellular DNA release in *Pseudomonas aeruginosa*. *PLoS one*, 2012; 7(10):e46718. [DOI:10.1371/journal.pone.0046718] [PMID:23056420]
12. Allen L, Dockrell DH, Pattery T, Lee DG, Cornelis P, Hellewell PG, Whyte MK. Pyocyanin production by *Pseudomonas aeruginosa* induces neutrophil apoptosis and impairs neutrophil-mediated host defenses in vivo. *J Immunol*. 2005; 174(6):3643-9. [DOI:<https://doi.org/10.4049/jimmunol.174.6.3643>] [PMID:15749902]
13. Lau GW, Hassett DJ, Ran H, Kong F. The role of pyocyanin in *Pseudomonas aeruginosa* infection. *Trends Mol Med*. 2004; 10(12):599-606. [DOI: 10.1016/j.molmed.2004.10.002] [PMID:15567330]
14. Usher LR, Lawson RA, Geary I, Taylor CJ, Bingle CD, Taylor GW, et al. Induction of neutrophil apoptosis by the *Pseudomonas aeruginosa* exotoxin pyocyanin: a potential mechanism of persistent infection. *J Immunol*. 2002; 168(4):1861-8. [DOI:<https://doi.org/10.4049/jimmunol.168.4.1861>] [PMID:11823520]

15. Bezanilla F. How membrane proteins sense voltage. *Nat Rev Mol Cell Biol.* 2008; 9(4):323-32. [DOI: 10.1038/nrm2376] [PMID 18354422]
16. Batovska DI, Todorova IT. Trends in utilization of the pharmacological potential of chalcones. *Curr Clin Pharmacol.* 2010; 5(1):1-29. [DOI:10.2174/157488410790410579] [PMID: 19891604]
17. Kabra, R, Chauhan N, Kumar A, Ingale P, Singh S. Efflux pumps and antimicrobial resistance: Paradoxical components in systems genomics. *Prog Biophys Mol Biol.* 2018. [DOI: 10.1016/j.pbiomolbio.2018.07.008] [PMID:30031023]
18. Jabeen I, Pleban K, Rinner U, Chiba P, Ecker GF. Structure-activity relationships, ligand efficiency, and lipophilic efficiency profiles of benzophenone-type inhibitors of the multidrug transporter P-glycoprotein. *J Med Chem.* 2012; 55(7):3261-73. [DOI:10.1021/jm201705f] [PMID:22452412]
19. Lowes DJ, Guiguemde WA, Connelly MC, Zhu F, Sigal MS, Clark JA, et al. Optimization of propafenone analogues as antimalarial leads. *J Med Chem.* 2011; 54(21):7477-85. [DOI: 10.1021/jm2005546] [PMID: 21955244]
20. Al Hussaini M, Hammouda EI, Hammouda AE. Optimizing pharmacotherapy of systemic lupus erythematosus: the pharmacist role. *Int J Clin Pharm.* 2014; 36(4):684-92. [DOI:10.1007/s11096-014-9966-1] [PMID:24986265]
21. Shao J, Feng G. Selective killing effect of oxytetracycline, propafenone and metamizole on A549 or HeLa cells. *Chin J Cancer Res.* 2013; 25(6):662. [DOI: 10.3978/j.issn.1000-9604.2013.11.05] [PMID:24385693]
22. Kouznetsova J, Sun W, Martínez-Romero C, Tawa G, Shinn P, Chen CZ, et al. Identification of 53 compounds that block Ebola virus-like particle entry via a repurposing screen of approved drugs. *Emerg Microbes Infect.* 2014; 3(1):1-7. [DOI:10.1038/emi.2014.88] [PMID:26038505]
23. Banach M, Piskorska B, Borowicz-Reutt KK. Propafenone enhances the anticonvulsant action of classical antiepileptic drugs in the mouse maximal electroshock model. *Pharmacol Rep.* 2016; 68(3):555-560. [DOI:10.1016/j.pharep.2016.01.002] [PMID:26894963]
24. Ngo ST, Fang ST, Huang SH, Chou CL, Huy PDQ, Li MS, Chen YC. Anti-arrhythmic medication propafenone a potential drug for Alzheimer's disease inhibiting aggregation of A β : in silico and in vitro studies. *J Chem Inf Model.* 2016; 56(7):1344-1356. [DOI:10.1021/acs.jcim.6b00029] [PMID:27304669]
25. Zheng WB, Li YJ, Wang Y, Yang J, Zheng CC, Huang XH, et al. Propafenone suppresses esophageal cancer proliferation through inducing mitochondrial dysfunction. *Am J Cancer Res.* 2017; 7(11):2245. [PMID:29218248]
26. Abonia R, Garay A, Castillo J, Insuasty B, Quiroga J, Nogueras M, et al. Design of Two Alternative Routes for the Synthesis of Naftifine and Analogues as Potential Antifungal Agents. *Molecules.* 2018; 23(3):520. [DOI:10.3390/molecules23030520] [PMID:29495412]
27. Chiba P, Burghofer S, Richter E, Tell B, Moser A, Ecker G. Synthesis, pharmacologic activity, and structure-activity relationships of a series of propafenone-related modulators of multidrug resistance. *J Med Chem.* 1995; 38(14):2789-93. [PMID:7629817]
28. Ivković B, Vladimirov S, Novaković R, Čupić V, Heinle H, Gojković-Bukarica L. The novel phenylpropiofenone derivatives induced relaxation of isolated rat aorta. *Arzneimittelforschung.* 2012; 62(07): 345-50. [DOI:10.1055/s-0032-1312617] [PMID:22628063]
29. Ivković B, Design, synthesis and biological activity of phenylpropiofenone aminoalcoxy derivatives [dissertation]. Faculty of Pharmacy, University of Belgrade, 2016

30. Knobloch JK, Horstkotte MA, Rohde H, Kaulfers PM, Mack D. Alcoholic ingredients in skin disinfectants increase biofilm expression of *Staphylococcus epidermidis*. *J Antimicrob Chemother.* 2002; 49(4):683-7. [PMID:11909845]
31. Basic J. Examination of correlation between chemical structure, physicochemical properties and retention parameters and antimicrobial activity of newly synthesized derivatives of propiophenone [dissertation]. Faculty of Pharmacy, University of Belgrade, 2016
32. Stepanović S, Vuković D, Hola V, Di Bonaventura G, Djukić S, Ćirković I, Ruzicka F. Quantification of biofilm microtiter plates: overview of testing conditions and practical recommendations for assessment of biofilm production by staphylococci. *APMIS.* 2007; 115:891-9. [DOI:10.1111/j.1600-0463.2007.apm_630.x] [PMID:17696944]
33. Glamočlija J, Ćirić A, Nikolić M, Fernandes Â, Barros L; Calheta RC, et al. Chemical characterization and biological activity of Chaga (*Inonotus obliquus*), a medicinal mushroom. *J Ethnopharmacol.* 2015; 162:323-32. [DOI: 10.1016/j.jep.2014.12.069] [PMID:25576897]
34. Sandy SM, Foong-Yee T. Anti-quorum sensing and antimicrobial activities of some traditional Chinese medicinal plants commonly used in South-East Asia. *Mal J Microbiol.* 2012; 8(1):11-20. [DOI:<http://dx.doi.org/10.21161/mjm.34911>]
35. SPSS Statistics. www.ibm.com/software/products/en/spss-statistics
36. Escaich S. Antivirulence as a new antibacterial approach for chemotherapy. *Curr Opin Chem Biol.* 2008;12(4):400-8. [DOI:10.1016/j.cbpa.2008.06.022] [PMID:18639647]
37. Lai S, Tremblay J, Déziel E. Swarming motility: a multicellular behavior conferring antimicrobial resistance. *Environ Microbiol.* 2009; 11(1):126-36. [DOI:10.1111/j.1462-2920.2008.01747.x] [PMID:18793317]
38. Morita Y, Tomida J, Kawamura Y. Responses of *Pseudomonas aeruginosa* to antimicrobials. *Front Microbiol.* 2014; 4:422. [DOI:10.3389/fmicb.2013.00422] [PMID:24409175]
39. Ramaswamy VK, Cacciotto P, Mallocci G, Ruggerone P, Vargiu AV. Multidrug efflux pumps and their inhibitors characterized by computational modeling. In: *Efflux-Mediated Antimicrobial Resistance in Bacteria*. Adis Cham. 2016; 797-831. [DOI: https://doi.org/10.1007/978-3-319-39658-3_30]
40. Li XZ, Nikaido H. Efflux-mediated drug resistance in bacteria: an update. *Drugs.* 2009; 69(12):1555. [DOI:10.2165/11317030-000000000-00000] [PMID:19678712]
41. Lima JLDC, Alves LR, Jacomé PRLDA., Neto B, Pacífico J, Maciel MAV et al. Biofilm production by clinical isolates of *Pseudomonas aeruginosa* and structural changes in LasR protein of isolates non biofilm-producing. *Braz J Infect Dis.* 2018; 22(2):129-36. [DOI:10.1016/j.bjid.2018.03.003] [PMID:29601791]
42. Ma L, Wang J, Wang S, Anderson EM, Lam JS, Parsek MR, Wozniak DJ. Synthesis of multiple *Pseudomonas aeruginosa* biofilm matrix exopolysaccharides is post-transcriptionally regulated. *Environ Microbiol.* 2012; 14(8):1995-2005. [DOI: 10.1111/j.1462-2920.2012.02753.x] [PMID:22513190]
43. Hay ID, Rehman ZU, Ghafoor A, Rehm BHA. Bacterial biosynthesis of alginates. *Chem Technol Biotechnol.* 2010; 85:752-9. [DOI:<https://doi.org/10.1002/jctb.2372>]
44. Britigan BE, Railsback MA, Cox CD. The *Pseudomonas aeruginosa* secretory product pyocyanin inactivates alpha1 protease inhibitor: implications for the pathogenesis of cystic fibrosis lung disease. *Infect Immun.* 1999; 67(3):1207-12. [PMID:10024562]

45. Rada B, Leto TL. Pyocyanin effects on respiratory epithelium: relevance in *Pseudomonas aeruginosa* airway infections. *Trends Microbiol.* 2013; 21(2):73-81. [DOI:10.1016/j.tim.2012.10.004] [PMID:23140890]
46. O'Malley YQ, Reszka KJ, Rasmussen GT, Abdalla MY, Denning GM, Britigan BE. The *Pseudomonas* secretory product pyocyanin inhibits catalase activity in human lung epithelial cells. *Am J Physiol Lung Cell Mol Physiol.* 2003; 285(5):L1077-L1086. [DOI:10.1152/ajplung.00198.2003] [PMID:12871859]
47. Look DC, Stoll LL, Romig SA, Humlicek A, Britigan BE, Denning GM. Pyocyanin and its precursor phenazine-1-carboxylic acid increase IL-8 and intercellular adhesion molecule-1 expression in human airway epithelial cells by oxidant-dependent mechanisms. *J Immunol.* 2005; 175(6):4017-23. [DOI:<https://doi.org/10.4049/jimmunol.175.6.4017>] [PMID:16148150]
48. Avdeef A, Strafford M, Block E, Balogh MP, Chambliss W, Khan I. Drug absorption in vitro model: filter-immobilized artificial membranes. 2. Studies of the permeability properties of lactones in *Piper methysticum* Forst. *Eur J Pharm Sci.* 2001; 14(4):271-80. [DOI:[https://doi.org/10.1016/S0928-0987\(01\)00191-9](https://doi.org/10.1016/S0928-0987(01)00191-9)] [PMID:11684401]

Table 1. *In vitro* effect of 5OF on the biofilm production of *Pseudomonas aeruginosa*

	5OF $\mu\text{g/ml}$					
	500	250	125	62.5	31.2	Positive control
<i>P. aeruginosa</i> Strain number	Category of biofilm production					
1	0	1	2	2	2	2
2	0	1	1	3	3	3
3	0	0	1	2	2	2
4	0	1	1	3	3	3
5	0	1	2	3	3	3
6	0	1	2	3	3	3
7	0	2	2	3	3	3
8	0	1	2	3	3	3
9	0	1	1	2	3	3
ATCC 27853	0	0	2	3	3	3

5OF: 3- (2-Fluoro-phenyl) -1- (2- (2-hydroxy-3-propylamino-propoxy) -phenyl) -propan-1-one hydrochloride; Positive control – bacterial growth in medium without tested compound; 0 – no biofilm production, 1 - weak biofilm production, 2 - moderate biofilm production, 3 - strong biofilm production.

Table 2. *In vitro* effect of 5CF3 on the biofilm production of *Pseudomonas aeruginosa*

	5CF3 μ g/ml					
	500	250	125	62.5	31.2	Positive control
<i>P. aeruginosa</i> Strain number	Category of biofilm production					
1	0	1	2	2	2	2
2	1	1	2	3	3	3
3	0	1	2	2	2	2
4	1	2	2	3	3	3
5	1	1	2	3	3	3
6	0	1	2	3	3	3
7	1	2	2	3	3	3
8	1	1	3	3	3	3
9	1	2	2	3	3	3
ATCC 27853	0	1	2	3	3	3

5CF3 :3- (2-Trifluoromethyl-phenyl) -1- (2- (2-hydroxy-3-propylamino-propoxy) -phenyl) -propan-1-one hydrochloride; Positive control – biofilm production in medium without tested compound; 0 – no biofilm production, 1 - weak biofilm production, 2 - moderate biofilm production, 3 - strong biofilm production.

Table 3. *In vitro* effect of *ortho*-fluorinated propafenone derivatives 5OF and 5CF3 on the production of pyocyanin in *Pseudomonas aeruginosa* strains

	5OF $\mu\text{g/ml}$		5CF3 $\mu\text{g/ml}$	
	500	250	500	250
<i>P. aeruginosa</i> Strain number	Pyocyanin production as% of positive control			
1	48.6	70.5	74.0	79.8
2	27.1	42.0	36.6	79.8
3	39.3	51.8	68.5	99.4
4	33.7	31.9	34.6	54.2
5	39.4	61.1	48.3	104.9
6	34.3	49.5	35.2	46.6
7	42.7	53.4	51.0	56.3
8	29.8	54.2	36.9	52.4
ATCC 27853	43.6	57.6	47.9	64.0

5OF: 3-(2-Fluoro-phenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride; 5CF3: 3-(2-Trifluoromethyl-phenyl)-1-(2-(2-hydroxy-3-propylamino-propoxy)-phenyl)-propan-1-one hydrochloride; Positive control – Pyocyanin production of each isolate in the absence of the tested compounds (100%).

Figure 1. *Pseudomonas aeruginosa* growth on Mueller-Hinton agar

