

## CASE REPORT / ПРИКАЗ БОЛЕСНИКА

# Hemolytic uremic syndrome complicating whooping cough

Vesna Stojanović<sup>1,2</sup>, Nenad Barišić<sup>1,2</sup>, Aleksandra Doronjski<sup>1,2</sup>, Dorottya Csuka<sup>3</sup>, Zoltán Prohászka<sup>3</sup><sup>1</sup>University of Novi Sad, Medical faculty, Novi Sad, Serbia;<sup>2</sup>Institute for Child and Youth Health Care of Vojvodina, Intensive Care Unit, Novi Sad, Serbia;<sup>3</sup>Semmelweis University, Research Laboratory and George Füst Complement Diagnostic Laboratory, 3rd Department of Medicine, Budapest, Hungary**SUMMARY**

**Introduction** We shall present a case of a two-month old infant who has developed a haemolytic uremic syndrome as an atypical complication of *Bordetella pertussis* infection. The observation that the development of haemolytic uremic syndrome is a late complication of *Bordetella pertussis* infection may be a clue for further studies.

**Case outline** A two-month-old female infant was admitted to the hospital because of fever, intensive cough, shortness of breath and poor feeding. Real-time polymerase chain reaction (PCR) for *Bordetella pertussis* was positive. A macrolide was introduced in therapy. On the eighth hospital day, the infant's condition improved, she became afebrile and eupneic. On the 16th hospital day, she developed signs of progressive respiratory distress and oliguric acute kidney injury. Hemolytic uremic syndrome (HUS) was diagnosed, so the therapy with the fresh frozen plasma (FFP) transfusion, therapeutic plasma exchange and peritoneal dialysis was initiated. Levels of ADAMTS13 (a disintegrin and metalloproteinase with a thrombospondin type 1 motif, member 13) were decreased, while the levels of factor H, factor B, and factor I were normal. Despite the full supportive and targeted care, severe multiple organ failure had developed and on the 24th hospital day the infant died.

**Conclusion** Further studies are necessary to identify the mechanism of potential interaction between pertussis toxins, pathophysiology of the infection and the interaction of complement activation, coagulation and the regulation of these cascades.

**Keywords:** hemolytic uremic syndrome; pertussis; infant

**INTRODUCTION**

Typical features of hemolytic uremic syndrome (HUS) are microangiopathic hemolytic anemia and acute thrombocytopenia, accompanied by acute kidney injury. In addition to infections with Shiga-like toxin-producing *Escherichia coli* (STEC-HUS), complement alternative pathway dysregulation may also predispose to atypical HUS (complement mediated aHUS). HUS may also develop in the context of severe infections, including for example *Streptococcus pneumoniae*, *Shigella dysenteriae* and *Bordetella pertussis*, various viruses, like H1N1 influenza, *Cytomegalovirus* and *Parvovirus B19* [1, 2, 3]. In addition to thrombotic thrombocytopenic purpura (TTP) and STEC-HUS, aHUS is classified in a group of thrombotic microangiopathies (TMA).

The incidence of aHUS and TTP is around 3/1,000,000 among children under 18. Patients with TTP have a good therapeutic response to plasma exchange (PE), whereas this is not the case in patients with aHUS. Mortality in patients with TTP is around 10%, and 25% in patients with aHUS during the acute phase [4, 5].

Despite the fact that vaccine is widely available, *Bordetella pertussis* infections are still very common. The classic symptoms of pertussis are

paroxysmal whooping cough, marked leukocytosis and related pulmonary leukostasis. So far, only four cases of HUS associated with *Bordetella pertussis* infection have been described in the literature [6–9].

**CASE REPORT**

A two-month-old female infant was admitted to the hospital due to fever, intensive cough, shortness of breath and poor feeding. First symptoms have begun two days before hospital admission.

Five days before admission, she received a combined five-component vaccine (diphtheria, tetanus, pertussis – acellular component, Polio-myelitis (Inactivated) Vaccine (Adsorbed) and Haemophilus Influenza Type B Conjugate Vaccine). She was the second child of the healthy non-consanguineous parents, born at term, weighting 2,900 g. Prenatal and perinatal period went without any complications.

On admission, she was conscious and alert, febrile, with normal vital parameters and normal body weight of 4,800 g. Lung auscultation revealed wheezing and bilateral, fine, late-inspiratory crackles on the bases of the lungs. All other physical findings were normal.

**Received • Примљено:**  
December 7, 2017

**Revised • Ревизија:**  
June 11, 2018

**Accepted • Прихваћено:**  
August 31, 2018

**Online first:** October 9, 2018

**Correspondence to:**

Vesna STOJANOVIĆ  
Faculty of Medicine  
University of Novi Sad  
Institute for Child and Youth  
Health Care of Vojvodina  
10 Hajduk Veljkova Street  
21000 Novi Sad, Serbia  
[vesna.stojanovic@mf.uns.ac.rs](mailto:vesna.stojanovic@mf.uns.ac.rs)

**Table 1.** Laboratory findings and clinical course

Parameter	Day 1	Day 5	Day 10	Day 14	Day 16	Day 17
Hemoglobin (g/l)	111	105	93	83	61	76
Platelet count (10 <sup>9</sup> /l)	552	778	1,137	481	95	46
White blood cell (10 <sup>9</sup> /l)	18	90	32	18	32	29
Creatinine (μmol/l)	/		/	/	328.5	398.47
CRP (mg/l)	45.9	36.7	negative	negative	13.34	/
Procalcitonine (ng/ml)	/	/	0.38	/	4.39	75.0
Clinical development	Coughing, heavy breathing	Tachydyspnoic, febrile, coughing, adynamic	Better overall condition, coughing	Paroxysmal whooping cough	Oliguria, edema, dyspnea	Anuria
Therapy	Antibiotic therapy (ceftazidime)	Antibiotic therapy (macrolide)	Antibiotic therapy (macrolide)	Antibiotic therapy (macrolide)	Intensive Care Unit	FFP, CAPD, TPE

FFP – frozen fresh plasma; CAPD – continuous ambulatory peritoneal dialysis, TPE – therapeutic plasma exchange

Initial laboratory findings are shown in Table 1. All other biochemical findings were within the normal ranges. Chest X-rays revealed pulmonary consolidation in perihilar and basal regions of the right lung and, also, regions of hyperinflation on both lungs. Ultrasound of the brain and abdomen showed normal findings.

The initial treatment included inhaled corticosteroids and bronchodilators and parenteral antibiotic therapy (ceftazidime). Despite the therapy, the child's condition worsened and she became tachydyspnoic, pale, adynamic and continuously febrile. On the fifth hospital day, marked leukocytosis was registered (Table 1). Peripheral blood smear showed prevalence of mature segmented granulocytes with toxic granulations, with normal red blood cell and platelet count and morphology.

Microbiologic and serologic tests confirmed the diagnosis of *Bordetella pertussis* infection (whooping cough). Real-time polymerase chain reaction (PCR) for *B. pertussis* was positive. Blood cultures were negative.

A macrolide was introduced in therapy. On the eighth hospital day, the infant's condition improved, she became afebrile and eupneic. This clinical melioration was accompanied with marked improvement of laboratory findings and reduction of WBC count.

On the 16th hospital day, the infant's condition suddenly worsened again. She had frequent attacks of heavy cough and developed signs of progressive respiratory distress with consequent oxyhemoglobin desaturation. She

became edematous (gained more than 600 g in weight in two days), with a decreased urine output (0.61 ml/kg/h), and hypertensive (BP 128/66 mmHg) so the infant was admitted to the Intensive Care Unit (ICU).

On admission to ICU, laboratory tests showed elevated procalcitonin levels, elevated WBC count, signs of hemolytic anemia and mild thrombocytopenia, as well as the signs of renal failure (Table 1). Peripheral blood smear revealed anisocytosis, polychromasia, presence of schistocytes and erythroblasts. Reticulocytes 23%, haptoglobin levels were < 0.10 g/l (0.3–2 g/l). Lactate dehydrogenase 64.15 μkat/l (< 7.52 μkat/l). Urea 15.8 mmol/L (1.8–6 mmol/l), creatinine 328.5 μmol/l (14–34 μmol/l), uric acid 1,311 μmol/l (65–319 μmol/l). Blood gas analysis showed decompensated metabolic acidosis. Hemostasis screening tests were normal.

Those clinical and laboratory findings raised suspicion of the presence of HUS, so we started the therapy with the fresh frozen plasma (FFP) transfusion. Hypertension was treated with calcium channel blocker.

Next day (17th hospital day), the infant became extremely tachydyspnoic, hence, she was intubated and mechanical ventilation was initiated. Because the child became anuric, the same day, Tenckhoff catheter for peritoneal dialysis (PD) was inserted and continuous peritoneal dialysis was immediately started. Initially, the dialysis solution with 2.3% glucose was used, and then after the edema was gone it was replaced with the solution with

**Table 2.** Findings of immunologic tests of parents and the neonate

Parameter (reference range)	Patient before FFP infusion (16th hospital day)	Patient after FFP infusion (17th hospital day)	Father	Mother
Classical pathway activity CH50/mL (48–103)	51	42		
C3 g/L (0.9–1.8)	0.88	0.54		
C1q mg/L (60–180)	36	40		
C4 g/L (0.15–0.55)	0.38	0.21	0.27	0.42
Alternative pathway activity % (70–105)	60	54		
Factor B % (70–130)	139	125	128	186
Factor H mg/L (250–880)	429	358	882	1209
Factor I % (70–130)	138	99	146	162
ADAMTS13 activity % (67–147)	43	50		

1.5% glucose. The initial fill volume of the dialysis solution was 10 ml/kg per exchange and was increasing slowly towards 20 ml/kg. Exchanges were done every 20–30 min. at the beginning, and after that hourly. Dwell time was 10 minutes, same as the drain time.

Also, the same day, two-lumen central venous catheter was inserted and the first session of therapeutic plasma exchange (TPE) was performed. During the hospitalization, a total of five TPE sessions have been done. Despite the applied therapy, signs of anuric renal injury, thrombocytopenia, anemia, and elevated lactate dehydrogenase levels continuously persisted.

Additional laboratory tests were carried out for the full work-up and TMAs differentiation. Findings of immunologic tests of the parents and the patient are shown in Table 2.

There was no available material for DNA extraction and genetic analysis of the patient. In both parents, complement profile was normal and the levels of some factors were elevated. Sequencing of the complete *CFH* gene of both parents was done, and no rare variations were observed, however, both parents turned out to carry the H3 aHUS risk haplotype in heterozygous manner.

Despite the full supportive and targeted care, severe multiple organ failure developed, and on the 24th hospital day the infant died.

## DISCUSSION

In the European cohort, 16% of aHUS cases were reported as a secondary HUS [10]. When a secondary HUS occurs, such as in the setting of different diseases, signs and symptoms of the primary disease can confound the diagnosis of aHUS [11].

Non-deficient ADAMTS13 activity (over 5–10%) supports the diagnosis of aHUS (under the condition that STEC test is negative). Of course, in these conditions, a differential diagnosis must be distinguished between aHUS and secondary HUS. Secondary HUS may be associated with different infections and sepsis, use of medications (anticancer molecules, immunotherapeutics, like cyclosporine, tacrolimus, and antiplatelet agents), malignancies and other underlying medical conditions such as autoimmune diseases, scleroderma and antiphospholipid syndrome. Familial aHUS occurs in about 10–20% of cases, the remaining patients having sporadic disease. Hypocomplementemia with a low level of C3 but normal C4 is a sign of alternative pathway dysregulation and activation, however, in 60–80% of the patients with aHUS levels of C3 are normal [12, 13, 2].

Due to the underlying immune-mediated, auto-inflammatory mechanisms and missing complement regulators, transfusions of FFP and plasmapheresis are therapeutic options for this syndrome. The aim of plasma therapy is to replace the mutant elements of the complement with normal elements in order to eliminate the pro-inflammatory and thrombogenic factors responsible for the symptoms. It is assumed that injections of fresh frozen plasma alone

are sufficient in the case of quantitative deficits [14]. But the long-term outcomes for this treatment are still not well-known [15].

In the case of the treatment failure (FFP, TPE), application of eculizumab (humanized monoclonal antibody against complement protein C5 terminal) is recommended [16, 17]. Studies that reported the benefits of these therapeutic choices in patients with aHUS are accumulating, but further evaluation is required to guide early and late therapeutic decisions like up-front treatment, treatment duration and discontinuation [18, 19, 20]. Baskin et al. [18] reported 10 paediatric patients with aHUS who did not respond to PE. Eculizumab improved their renal function and quality of life. Greenbaum et al. [19] prospectively evaluated efficacy and safety of eculizumab in paediatric patients with aHUS. Their findings establish the efficacy and safety of eculizumab for the paediatric patients with aHUS and the recommendation is to start the therapy with it as soon as possible after establishing the aHUS diagnosis. Considering the cost of the treatment, the cost/benefit of this cure should be estimated more thoroughly in the future.

This drug was neither registered nor available in Serbia, at the time of our patient's treatment.

*B. pertussis* produces a number of virulence factors that are involved in the pathogenesis and manifestations of the disease. Toxins of *B. pertussis* and other virulence factors enable adhesion of bacteria, locally injure epithelium, cause leukocyte dysfunction and macrophage cytotoxicity, increase release of proapoptotic and pro-inflammatory cytokines such as TNF-alpha and IL-6. In addition to the damage to the epithelium, these factors injure the endothelium, which causes a pro-coagulant pathway activation [21].

The infant presented in our report has received the first dose of pertussis vaccine two days prior getting the disease, so she has not developed the immunity.

So far, only four cases of HUS associated with *Bordetella pertussis* infection have been described in the literature [6–9]. The association of *Bordetella pertussis* infection and HUS was first described by Berners et al. [6] in a newborn with complement factor H (*CFH*) mutation, which was treated with peritoneal dialysis and ended fatally. In this patient, an abnormal band was identified on Factor H Western-blot, indicative for the presence of an abnormal complement regulator. After that, Pela et al. [9] described the case of a 42-day-old infant treated with hemodiafiltration and FFP infusions. In this case mutations in the genes encoding factor H and MCP were not detected. This patient survived with a completely recovered renal function. Chaturvedi et al. [7] described the case of a 28-day-old baby who was treated with FFP infusions and did not require dialysis. Similarly to previously mentioned case, mutations in the genes encoding factor H, factor I, factor B and ADAMTS13 were not identified. The patient survived, fully recovered and maintained a normal renal function. The most recent case was reported by Cohen-Ganea et al. [8]. In this case, complement (C3, C4, CH50) and factor H levels were within the normal range.

Our patient was a two-month-old infant who had developed HUS three weeks after starting the whooping cough, and who was treated by peritoneal dialysis and TPE without a favorable response. Death of the patient was caused by HUS and severe sepsis which resulted in multiple organ dysfunction.

In the acute phase, very low C1q levels with decreased C3 concentration and combined (classical and alternative pathway) complement consumption were observed. We apprehended that as a result of ongoing severe infection, not as dysregulation of the alternative pathway. In line with this, there was no evidence of Factor H mutation in this patient, since the parents did not carry rare variations in *CFH*. Hence, we consider the possibility of a cause-effect relationship between *B. pertussis* infection and HUS in our patient. *B. pertussis* infection may be capable to stimulate the inflammatory cells to release the cytokines which determine microangiopathy and HUS. No abnormalities of factor H were found in the parents of our patient, however, due to the lack of available samples for DNA extraction we were unable to completely sequence all of the important genes of complement regulators and factors. Therefore, we can only assume that *B. pertussis* may cause HUS in the children without any predisposing factors. The infant developed HUS in the later phase of the infection after a brief period of clinical improvement (from the eighth to

the 14th hospital day). From this aspect our patient was similar to the three previously reported patients who had HUS in the context of an infection caused by *B. pertussis*. These patients also developed a severe HUS after a period of clinical improvement – in the period between the sixth and 42nd hospital day [6, 7, 9]. The patient described by Cohen-Ganelin et al. [8] was in very difficult condition with paroxysmal coughing and extreme excitability, all the time until the moment of HUS diagnosis (the 10th hospital day). So far, all described cases have developed HUS between the 12th and 45th hospital day since the start of the illness (first symptoms of a whooping cough).

Further studies are necessary to identify the mechanism of a potential interaction between pertussis toxins, pathophysiology of the infection and the interaction of complement activation, coagulation and the regulation of these cascades. The observation of the development of HUS as a late complication of *B. pertussis* infection may be a clue for further studies.

## ACKNOWLEDGEMENT

The technical assistance of Ágnes Szilágyi, Márta Kókai, Éva Szendrei, Beáta Takács and Edina Szabó is acknowledged with many thanks.

## REFERENCES

- Besbas N, Karpman D, Landau D, Loirat C, Proesmans W, Remuzzi G, et al. A classification of hemolytic uremic syndrome and thrombotic thrombocytopenic purpura and related disorders. *Kidney Int.* 2006; 70(3):423–31.
- Salvadori M, Bertoni E. Update on haemolytic uremic syndrome: diagnostic and therapeutic recommendations. *World J Nephrol.* 2013; 2(3):56–76.
- Siegler R, Oakes R. Hemolytic uremic syndrome; pathogenesis, treatment, and outcome. *Curr Opin Pediatr.* 2005; 17(2):200–4.
- Ariceta G, Besbas N, Johnson S, Karpman D, Landau D, Licht C. European Paediatric Study Group for HUS. Guideline for the investigations and initial therapy of diarrhoea-negative haemolytic uremic syndrome. *Pediatr nephrol.* 2009; 24(4):687–96.
- Noris M, Remuzzi G. Atypical haemolytic uremic syndrome. *N Engl J Med.* 2009; 361:1676–87.
- Berner R, Krause MF, Gordjani N, Zipfel PF, Boehm N, Krueger M, et al. Haemolytic uremic syndrome due to an altered factor H triggered by neonatal pertussis. *Pediatr Nephrol.* 2002; 17(3):190–2.
- Chaturvedi S, Licht C, Langlois V. Haemolytic uremic syndrome caused by *Bordetella pertussis* infection. *Pediatr Nephrol.* 2010; 25(7):1361–4.
- Cohen GE, Davidovits M, Amir J, Prais D. Severe *Bordetella pertussis* infection associated with hemolytic uremic syndrome. *Isr Med Assoc J.* 2012; 14(7):456–8.
- Pela I, Seracini D, Caprioli A, Castelletti F, Giammanco A. Hemolytic uremic syndrome in an infant following *Bordetella pertussis* infection. *Eur J Clin Microbiol Infect Dis.* 2006; 25(8):515–7.
- Noris M, Caprioli J, Bresin E, Mossali C, Pianetti G, Gamba S, et al. Relative role of genetic complement abnormalities in sporadic and familial aHUS and their impact on clinical phenotype. *Clin J Am Soc Nephrol.* 2010; 5(10):1844–59.
- Nester CM, Thomas CP. Atypical hemolytic uremic syndrome: what is it, how is it diagnosed, and how is it treated? *Hematology.* 2012; 2012:617–25.
- Laurence J. Atypical hemolytic uremic syndrome (aHUS): making the diagnosis. *Clin Adv Hematol Oncol.* 2012; 10(Suppl 17):1–12.
- Noris M, Mescia F, Remuzzi G. STEC-HUS, atypical HUS and TTP are all diseases of complement activation. *Nat Rev Nephrol.* 2012; 8(11):622–33.
- Loirat C, Garnier A, Sellier-Leclerc AL, Kwon T. Plasmatherapy in atypical hemolytic and uremic syndrome. *Semin Thromb Hemost.* 2010; 36(6):673–81.
- Lapeyraque AL, Wagner E, Phan V, Clermont MJ, Merouani A, Frémeaux-Bacchi V, et al. Efficacy of plasma therapy in atypical hemolytic uremic syndrome with complement factor H mutations. *Pediatr Nephrol.* 2008; 23(8):1363–6.
- Magen D, Oliven A, Shechter Y, Elhasid R, Joseph GB, Zelikovic I. Plasmapheresis in a very young infant with atypical haemolytic uremic syndrome. *Pediatr Nephrol.* 2001; 16(1):87–90.
- Szarvas N, Szilagy A, Tasic V, Nushi-Stavileci V, Sofijanov A, Gucev Z, et al. First-line therapy in atypical hemolytic uremic syndrome: consideration on infants with a poor prognosis. *Ital J Pediatr.* 2014; 40(1):101.
- Baskin E, Gulleroglu K, Kantar A, Bayrakci U, Ozkaya O. Success of eculizumab in the treatment of atypical haemolytic uremic syndrome. *Pediatr Nephrol.* 2015; 30(5):783–9.
- Greenbaum LA, Fila M, Ardissino G, Al-Akash SI, Evans J, Henning P, et al. Eculizumab is a safe and effective treatment in pediatric patients with atypical hemolytic uremic syndrome. *Kidney Int.* 2016; 89(3):701–11.
- Picard C, Burtet S, Bornet C, Curti C, Montana M, Vanella P. Pathophysiology and treatment of typical and atypical hemolytic uremic syndrome. *Pathol Biol.* 2015; 63(3):136–43.
- Heininger U. Recent progress in clinical and basic pertussis research. *Eur J Pediatr.* 2001; 160:203–13.

## Хемолитичко-уремички синдром као компликација великог кашља

Весна Стојановић<sup>1,2</sup>, Ненад Баришић<sup>1,2</sup>, Александра Дороњски<sup>1,2</sup>, Доротија Чука<sup>3</sup>, Золтан Прохаска<sup>3</sup>

<sup>1</sup>Универзитет у Новом Саду, Медицински факултет, Нови Сад, Србија;

<sup>2</sup>Институт за здравствену заштиту деце и омладине Војводине, Одељење интензивне неге и терапије, Нови Сад, Србија;

<sup>3</sup>Универзитет *Semmelweis*, Истраживачка лабораторија и додатна дијагностичка лабораторија *George Füst*, Треће одељење медицине, Будимпешта, Мађарска

### САЖЕТАК

**Увод** Приказан је случај двомесечног одојчета које је развило хемолитичко-уремички синдром као атипичну компликацију инфекције бактеријом *Bordetella pertussis*. Чињеница да је развој хемолитичко-уремичког синдрома касна компликација инфекције бактеријом *Bordetella pertussis* може бити полазна тачка за планирање даљих студија.

**Приказ болесника** Двомесечно женско одојче је хоспитализовано на Клиници за педијатрију због повишене температуре, интензивног кашља, диспнеје и одбијања хране. Ланчана реакција полимеразом у реалном времену на бактерију *Bordetella pertussis* је била позитивна. Започета је терапија макролидима и осмог дана хоспитализације клиничко стање детета се побољшало, постало је афебрилно и еупноично. Шеснаестог дана хоспитализације дете је развило клиничке

знаке прогресивног респираторног дистреса и олигуричног акутног бубрежног оштећења. Дијагностикован је хемолитичко-уремички синдром, те је започета терапија трансфузијама свеже смрзнуте плазме, а затим и терапија изменама плазме и перитонеумска дијализа. Нивои металопротеиназе ADAMTS13 били су снижени, док су нивои фактора *H*, *B* и *I* били нормални. Упркос спроведеној терапији, у даљем току развија се мултиорганска дисфункција и 24. дана хоспитализације долази до смртог исхода.

**Закључак** Потребне су додатне студије да би се утврдили механизми могуће интерреакције између токсина пертусиса, патофизиологије инфекције и интеракције активације комплемената, коагулације и регулације ових каскада.

**Кључне речи:** хемолитичко-уремички синдром; велики кашаљ; одојче